

Carriage of methicillin-resistant *Staphylococcus aureus* in a Queensland Indigenous community

Susan Vlack, Leonie Cox, Anton Y Peleg, Condy Canuto, Christine Stewart, Alzira Conlon, Alex Stephens, Philip Giffard, Flavia Huygens, Adam Mollinger, Renu Vohra and James S McCarthy

Community-acquired infections with *Staphylococcus aureus* have, until very recently, been reliably treated with β -lactam antibiotics. This is in contrast to hospital-acquired infections, where for more than 20 years methicillin-resistant *S. aureus* (MRSA) has been a problem. In the late 1980s, a strain of MRSA arose spontaneously in remote Aboriginal communities of Western Australia.^{1,2} This strain, termed WA-MRSA,¹ has since become established in western and northern Australia, in both the hospital and the community setting. More recently, community-acquired methicillin-resistant *S. aureus* (CA-MRSA) has emerged along the eastern seaboard of Australia, with the South West Pacific strain, referred to as Sequence Type 30 (ST30), predominantly affecting individuals of Polynesian descent, and the Queensland clone (ST93) affecting white people.^{3,4}

Although CA-MRSA has predominantly caused skin and soft-tissue infections, invasive life-threatening infection is now being reported, including necrotising pneumonia, septic shock and death.⁵ The increased virulence of this organism has been associated with a specific virulence factor named Panton–Valentine leukocidin (PVL), a bacterial toxin that can mediate leukocyte and tissue destruction.⁶ The combination of this enhanced virulence with resistance to the most commonly prescribed antistaphylococcal antibiotics makes this organism an important public health threat.

To date, the prevalence of CA-MRSA in Australia has not been formally assessed by a community-based study. Isolates have been obtained from patients requiring hospital admission, without study of the community prevalence.^{1–5,7} Collection of prevalence data is critical for formulating health care policies pertaining to empiric antibiotic use, particularly for suspected staphylococcal infections.

Following a death in our hospital caused by CA-MRSA infection in a young adult, as well as a number of serious CA-MRSA infections with epidemiological links to an Indigenous community,⁵ we undertook a study to assess the prevalence of CA-MRSA carriage in this community.

ABSTRACT

Objective: To determine the prevalence of community-acquired methicillin-resistant *Staphylococcus aureus* (CA-MRSA) carriage and infection among children living in an Indigenous community in Queensland.

Design, setting and participants: Swabs for culture of *S. aureus* were collected from the nose, throat and skin wounds of primary school children.

Main outcome measures: MRSA carriage, antibiotic sensitivity, genotype, and presence of the virulence factor Panton–Valentine leukocidin (PVL); and epidemiological risk factors for MRSA carriage.

Results: 92 (59%) of 157 eligible children were included in the study. Twenty-seven (29%) carried *S. aureus*; 14 of these (15% of total) carried MRSA. MRSA was isolated from 29% of wound swabs, 8% of nose swabs, and 1% of throat swabs. Fourteen of 15 MRSA isolates were sensitive to all non- β -lactam antibiotics tested. Eight children (9%) carried CA-MRSA clonal types: six carried the Queensland clone (ST93), and two carried the South West Pacific clone (ST30). All these isolates carried the virulence factor PVL. The remaining six children carried a hospital-associated MRSA strain (ST5), negative for PVL.

Conclusions: We have identified a high prevalence of CA-MRSA carriage in school children from a Queensland Indigenous community. In this setting, antibiotics with activity against CA-MRSA should be considered for empiric therapy of suspected staphylococcal infection. Larger community-based studies are needed to improve our understanding of the epidemiology of CA-MRSA, and to assist in the development of therapeutic guidelines for this important infection.

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METHODS

Study population

The study was conducted in an Indigenous community in south-east Queensland with a population of about 2000. In collaboration with local Indigenous community representatives, the local primary school was used as the site for data and sample collection. Most of the town's children are enrolled at this school. All children from grades one to seven were eligible for the study, which took place over 3 days in October 2004.

Data collection

After written informed consent had been obtained from parents or guardians, a written questionnaire was completed by parents/guardians to collect demographic and relevant risk factor data. Swabs from both anterior nares and the throat were collected from the participating children. A skin swab was collected from any skin lesions evident on exposed skin. If obvious skin infection was identified, the children and

their parents were referred to local health service providers.

Microbiology

All swabs were transported to a central laboratory in Brisbane, and processed within 12 hours. Bacterial isolates were identified and antibiotic sensitivity determined using standard laboratory methods.⁸ Susceptibility to clindamycin was extrapolated from erythromycin susceptibility unless erythromycin resistance was observed, in which case inducible clindamycin was tested for. Culture and sensitivity results were made available to local health care providers, and parents or carers were advised by letter of the results and recommended action.

Genotyping and PVL detection

Genotyping was performed on all MRSA isolates using a recently developed method that uses seven single nucleotide polymorphisms derived from the multilocus sequence typing database. The presence of the virulence factor PVL and the *mecA* gene

1 Consultative research process

- One researcher familiar with community processes introduced the team to key stakeholders in health and education.
- A research implementation plan was developed and approved locally with community representatives, health and education staff and researchers via the established Health Action Group.
- An ethics proposal was submitted and approved.
- Support was gained from the local school's headmaster, and a staff forum and information session was conducted at the school by project staff.
- Information, consent and survey forms were distributed to all parents, with the assistance of the school, and were then collected back at school.
- The school agreed that sample collection could be conducted on the school premises, that school staff would accompany the children to the site where samples were collected, and that staff would assist by verifying that the consent form matched the child in attendance.
- Funds were provided by Queensland Health to pay a local resident to work on the project and to pay a local artist for graphics that were used on skin health and project promotional material (Box 2).
- The project was implemented in a community-recommended time frame.
- A skin infection awareness campaign was undertaken by a local community member. This involved radio broadcasts, posters, meetings, participation in a community event, group discussions and distribution of printed information through various networks. Information about the project and advice on responding to skin infections and maintaining healthy skin were included. Swabs were then collected.
- The project provided field experience for a student (Bachelor of Applied Health Sciences [Indigenous Health]) working with a local project officer, and an elective placement for a medical student.
- Initial study results were returned to the community committee and doctors.
- A final draft report was returned to the community for their perusal and approval.
- A radio broadcast and items in school and community newsletters provided general feedback of study findings.
- The community committee was given an opportunity to view this article before publication.
- Opportunity is provided for further community-driven skin health research. ♦

was determined by polymerase chain reaction (PCR).^{9,10}

Statistical analysis

Statistical data entry and analysis were performed using SPSS version 13 (SPSS Inc, Chicago, Ill, USA). Statistical comparison of groups was performed using Pearson's χ^2 or Fisher's exact test. Continuous data were evaluated using the Mann-Whitney rank sum test. All *P* values were 2-tailed, and *P* < 0.05 was considered significant.

Ethics

The project proposal was approved by the community Health Action Group, in keeping with research protocols involving Aboriginal communities. The relevant Queensland Health District Manager and local school principal also gave written support. Approval was obtained from the University of Queensland Ethics Committee. Research staff external to the community worked closely with local staff during all project activities, and a local person was employed as a project officer, as advised by the Health Action Group. The consultative research process is summarised in Box 1.

RESULTS

Ninety-two (59%) of 157 eligible children were included. The remaining children were either unavailable during the study period (*n* = 2) or consent was not obtained (*n* = 63). The median age of the study participants was 8 years (range, 5–13 years); 63 (68%) were girls. Each school grade was represented, contributing between 8% and 18% of the study group. Most of the children (93%) had lived in the community for more than 1 year. The 92 children lived in 51 households containing three to 14 people, with 56% having six to 10 residents and 15% having 11 or more. Most children (87%) lived with smokers. Some children (13%) had received antibiotics in the week before swab collection.

Nose swabs were collected from all study participants, and all except one had a throat swab. Skin lesions were observed on 24 (26%) children, and swabs were collected from these lesions. Fourteen children with lesions (15% of the study group) required referral to local health providers.

Culture results from different swab sites are shown in Box 3. Of the 92 children, 27 (29%) were colonised or infected with *S. aureus*, including 14 carrying MRSA (15% overall prevalence); isolates from the other

13 children were methicillin-sensitive. All but one of the MRSA isolates were sensitive to all non- β -lactam antibiotics tested. One isolate was resistant to erythromycin and had inducible resistance to clindamycin. In contrast to other studies undertaken in northern Australia, we observed a low rate of carriage of Group A streptococci (one throat, one skin sore).

Three MRSA clonal types were found among the 14 children carrying MRSA (Box 4). Eight children (9%) carried CA-MRSA clonal types: six had the Queensland clone (ST93), and two had the South West Pacific clone (ST30). All these isolates were positive for the virulence factor PVL. The other six children carried a strain previously observed in hospitals (ST5), and all these isolates were negative for PVL. Of note, this strain is distinct from isolates commonly found in Australian hospital settings. ST5 and close relatives are frequently isolated in health care facilities in many parts of the world. However, in Australia, strains of this lineage are isolated from health care facilities much less frequently than other clones, such as ST239.

Of the 24 children from whom skin swabs were collected (in addition to nose and throat swabs), nine (38%) had MRSA; seven of these isolates (78%) were CA-MRSA clonal types. Of these seven CA-MRSA isolates, six were cultured from the skin only and one from the skin and nose.

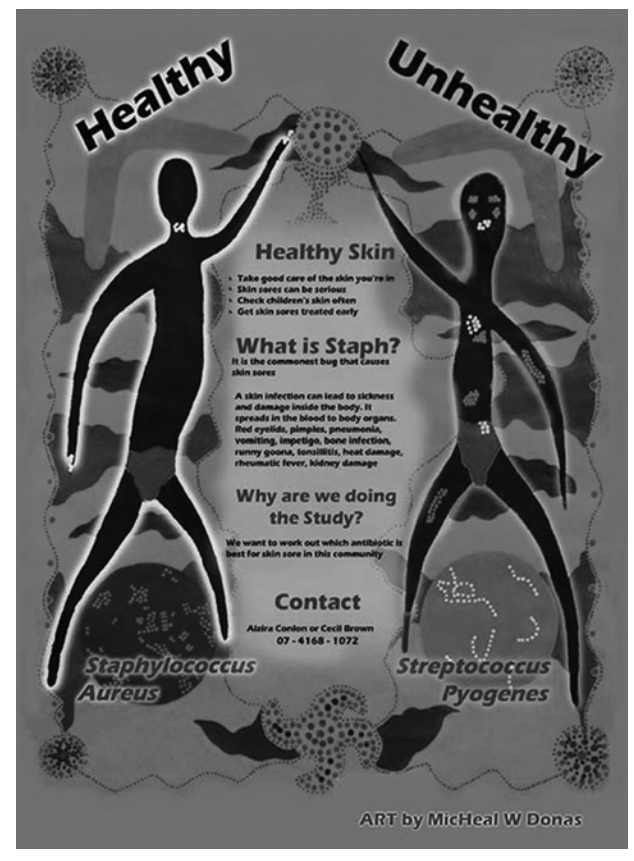
DISCUSSION

CA-MRSA has emerged as an important pathogen with the potential to cause life-threatening infections. To date, there have been few population-based studies of CA-MRSA prevalence, and even fewer that have included molecular typing. In this study, we found the community prevalence of MRSA in school children from a Queensland Indigenous population to be 15%, with more than half of these children carrying PVL-positive CA-MRSA strains (9% prevalence). Although the overall *S. aureus* carriage rate in our study (29%) was similar to that reported by others,^{11,12} the proportion of participants carrying CA-MRSA was higher than reported in many studies, where prevalence has ranged from 0.2%¹² to 2.5%.¹³

We believe our results warrant a change in empiric antibiotic prescribing in communities that have such high prevalence of CA-MRSA (9%), particularly for patients presenting with life-threatening infections such as severe pneumonia or undifferentiated

2 Poster developed for promoting the screening project

Artwork by Michael Donas.



3 Results of swab culture for *Staphylococcus aureus*

Site	Methicillin-sensitive isolates	Methicillin-resistant isolates	Number of subjects with isolates from this site only
Nose (n = 92)	11 (61%)	7 (28%)	14 (15%)
Throat (n = 91)	1 (33%)	1 (33%)	1 (1%)
Skin (n = 24)	5 (36%)	7 (50%)	7 (29%)

4 Methicillin-resistant *Staphylococcus aureus* (MRSA) genotypes isolated

MRSA sequence type	Number of isolates	Isolate site			Presence of PVL virulence factor
		Nose	Throat	Skin	
5	6	5	1	0	0/6
30	3	1	0	2	3/3
93	6	1	0	5	6/6

PVL = Pantón-Valentine leukocidin. ♦

ated sepsis. For these conditions, we advocate including vancomycin in empiric therapy while cultures are pending. More commonly, patients present with skin and soft-tissue infections that often respond to drainage alone, but oral antibiotic therapy may be required.

A characteristic feature of CA-MRSA, as observed in our study, is sensitivity to many non- β -lactam antibiotics with useful antistaphylococcal activity, such as clindamycin and trimethoprim-sulfamethoxazole. Of importance, if erythromycin resistance is present, clinicians should avoid clindamycin therapy unless the laboratory has tested for inducible clindamycin resistance. Given the changing epidemiology of *S. aureus* infections and antibiotic susceptibility in the community, appropriate clinical specimens for culture and sensitivity testing should be collected where antimicrobial therapy is indicated. Other interventions to reduce the prevalence of skin sepsis require consideration. These include public health programs to control scabies¹⁴ and the provision of swimming pools.¹⁵

isolates were from the skin only. The explanation for this site predilection is unclear, but may relate to strain-specific characteristics such as adherence factors. Of note, most children carrying CA-MRSA had some type of wound on exposed skin. In previous studies, skin and soft-tissue infection has been shown to be a risk factor for CA-MRSA carriage.^{12,16} This association is best explained by the presence of PVL, which is important in the pathogenesis of skin and soft-tissue infections. Our observations suggest that screening using nasal swabs alone may be inadequate for detecting CA-MRSA.¹⁷

Limitations of our study include the small sample size and possible selection bias due to differences in carriage between children whose parents agreed to participate and those who did not. Males, who generally have higher rates of carriage of *S. aureus*, were under-represented. The study was not sufficiently powered to investigate risk factors for acquisition, including household size. Household crowding remains an important and unresolved issue in many

Indigenous communities throughout Australia.^{18,19} Our results relate to a single community and may not be generalisable. However, they do suggest that such organisms may be prevalent in a much wider geographical area. Ongoing and larger community-based studies are needed to improve our understanding of the epidemiology of CA-MRSA. Such studies will guide the formulation of antibiotic policies and the development of preventive strategies for this important infection.

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COMPETING INTERESTS

None identified.

AUTHOR DETAILS

Susan Vlack, FAFPHM, Lecturer¹
 Leonie Cox, PhD, Lecturer¹
 Anton Y Peleg, FRACP, Infectious Diseases Registrar²
 Condyl Canuto, MAE, Lecturer¹
 Christine Stewart, Centre Manager³
 Alzira Conlon, Health Worker³
 Alex Stephens, BSc(Hons), PhD Student⁴
 Philip Giffard, PhD, Associate Professor⁴
 Flavia Huygens, PhD, Research Fellow⁴
 Adam Mollinger, MB BS, Medical Student¹
 Renu Vohra, FRCPA, Clinical Microbiologist⁶

RESEARCH

James S McCarthy, FRACP, Associate Professor of Tropical Medicine and Infectious Diseases^{1,6}

1 School of Population Health, University of Queensland, Brisbane, QLD.

2 Royal Melbourne Hospital, Melbourne, VIC.

3 Queensland Health Community Health Service, QLD.

4 Cooperative Research Centre for Diagnostics, Queensland University of Technology, Brisbane, QLD.

5 Queensland Medical Laboratory, Brisbane, QLD.

6 Queensland Institute of Medical Research, Brisbane, QLD.

Correspondence: j.mccarthy@uq.edu.au

REFERENCES

- 1 Riley TV, Pearman JW, Rouse IL. Changing epidemiology of methicillin-resistant *Staphylococcus aureus* in Western Australia. *Med J Aust* 1995; 163: 412-414.
- 2 Maguire GP, Arthur AD, Boustead PJ, et al. Clinical experience and outcomes of community-acquired and nosocomial methicillin-resistant *Staphylococcus aureus* in a northern Australian hospital. *J Hosp Infect* 1998; 38: 273-281.
- 3 Munckhof WJ, Schooneveldt J, Coombs GW, et al. Emergence of community-acquired methicillin-resistant *Staphylococcus aureus* (MRSA) infection in Queensland, Australia. *Int J Infect Dis* 2003; 7: 259-264.
- 4 Nimmo GR, Schooneveldt J, O'Kane G, et al. Community acquisition of gentamicin-sensitive methicillin-resistant *Staphylococcus aureus* in southeast Queensland, Australia. *J Clin Microbiol* 2000; 38: 3926-3931.
- 5 Peleg AY, Munckhof WJ, Kleinschmidt SL, et al. Life-threatening community-acquired methicillin-resistant *Staphylococcus aureus* infection in Australia. *Eur J Clin Microbiol Infect Dis* 2005; 24: 384-387.
- 6 Zetola N, Francis JS, Nuermberger EL, et al. Community-acquired methicillin-resistant *Staphylococcus aureus*: an emerging threat. *Lancet Infect Dis* 2005; 5: 275-286.
- 7 Nimmo GR, Coombs GW, Pearson JC; Australian Group for Antimicrobial Resistance (AGAR). Methicillin-resistant *Staphylococcus aureus* in the Australian community: an evolving epidemic. *Med J Aust* 2006; 184: 384-388.
- 8 Bell SM, Gatus BJ, Pham JN, Rafferty DL. Antibiotic susceptibility testing by the CDS method. A manual for medical and veterinary laboratories. 3rd ed. Sydney: South Eastern Area Laboratory Services, 2004.
- 9 Robertson GA, Thiruvankataswamy V, Shilling H, et al. Identification and interrogation of highly informative single nucleotide polymorphism sets defined by bacterial multilocus sequence typing databases. *J Med Microbiol* 2004; 53: 35-45.
- 10 Stephens AJ, Huygens F, Inman-Bamber J, et al. Methicillin-resistant *Staphylococcus aureus* genotyping using a small set of polymorphisms. *J Med Microbiol* 2006; 55: 43-51.
- 11 Bogaert D, van Belkum A, Sluijter M, et al. Colonisation by *Streptococcus pneumoniae* and *Staphylococcus aureus* in healthy children. *Lancet* 2004; 363: 1871-1872.
- 12 Leman R, Alvarado-Ramy F, Pocock S, et al. Nasal carriage of methicillin-resistant *Staphylococcus aureus* in an American Indian population. *Infect Control Hosp Epidemiol* 2004; 25: 121-125.
- 13 Hussain FM, Boyle-Vavra S, Daum RS. Community-acquired methicillin-resistant *Staphylococcus aureus* colonization in healthy children attending an outpatient pediatric clinic. *Pediatr Infect Dis J* 2001; 20: 763-767.
- 14 Wong LC, Amega B, Connors C, et al. Outcome of an interventional program for scabies in an Indigenous community. *Med J Aust* 2001; 175: 367-370.
- 15 Lehmann D, Tennant MT, Silva DT, et al. Benefits of swimming pools in two remote Aboriginal communities in Western Australia: intervention study. *BMJ* 2003; 327: 415-419.
- 16 Hidron AI, Kourbatova EV, Halvosa JS, et al. Risk factors for colonization with methicillin-resistant *Staphylococcus aureus* (MRSA) in patients admitted to an urban hospital: emergence of community-associated MRSA nasal carriage. *Clin Infect Dis* 2005; 41: 159-166.
- 17 Moellering RC. The growing menace of community-acquired methicillin-resistant *Staphylococcus aureus*. *Ann Intern Med* 2006; 144: 368-370.
- 18 Building a better future: Indigenous housing to 2010. Housing Minister's Conference, 4 May 2001. Available at: http://www.facs.gov.au/internet/facsinternet.nsf/aboutfacs/programs/community-indig_housing_2010.htm (accessed Apr 2006).
- 19 House of Representatives Standing Committee on Family and Community Affairs. Health is life. Report on the inquiry into Indigenous health. Canberra: Commonwealth of Australia, 2000. Available at: <http://www.aph.gov.au/house/committee/fca/indhea/inqinde2.htm> (accessed Apr 2006).

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